

# How should functional relationships be evaluated using phylogenetic comparative methods? A case study using metabolic rate and body temperature

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1 **Running Head:** Functional relationships and PCMs

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## 6 **Conflicts of Interest**

7 The authors have no conflicts of interest to declare.

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9 Supplementary Material and complete scripts and data are available on Dryad, <https://doi.org/10.5061/dryad.z612jm6bj>.

<sup>10</sup> **Author contributions**

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## Abstract

Phylogenetic comparative methods are often used to test functional relationships between traits. However, million-year macroevolutionary observational datasets cannot definitively prove causal links between traits — correlation does not equal causation and experimental manipulation over such timescales is impossible. While this caveat is widely understood, it is less appreciated that different phylogenetic approaches imply different causal assumptions about the functional relationships of traits. In order to make meaningful inferences, it is critical that our statistical methods make biologically reasonable assumptions. Here we illustrate the importance of causal reasoning in comparative biology by examining a recent study by [Avaria-Llautureo et al. \(2019\)](#) that tested for the evolutionary coupling of metabolic rate and body temperature across endotherms and found that these traits were unlinked through evolutionary time and that body temperatures were, on average, higher in the early Cenozoic than they are today. We argue that the causal assumptions embedded into their models made it impossible for them to test the relevant functional and evolutionary hypotheses. We re-analyze their data using more biologically appropriate models and find support for the exact opposite conclusions, corroborating previous evidence from physiology and paleontology. We highlight the vital need for causal thinking, even when experiments are impossible.

## Introduction

The evolutionary causes and consequences of endothermy have fascinated biologists for decades. But this is naturally a difficult thing to study: the evolutionary events we are interested in occurred over the course of hundreds of millions of years. At this scale, direct tests of causal hypotheses are impossible. Nonetheless by piecing together multiple lines of evidence, including physiological experiments, mathematical theory, and the paleontological record, we have gained a rich understanding of the relationships between metabolic rates, body mass, and temperature and how they might have evolved ([Grigg et al. 2004](#); [Clarke et al. 2010](#); [Lovegrove 2017](#)), even as mysteries remain.

Macroevolutionary comparative analyses complement these approaches; by leveraging interspecific data in a phylogenetic framework, we can potentially gain new insights into how physiological processes ([White et al. 2009](#); [Uyeda et al. 2017](#); [White et al. 2019](#)) and scaling relationships, more generally, evolve ([Hansen and Bartoszek 2012](#); [Pélabon et al. 2014](#); [Voje et al. 2014](#)). And while developments in this area are

41 tremendously exciting, it is critical to keep in mind that inferences drawn from phylogenetic statistical meth-  
42 ods, just like many other statistical methods, have embedded, and often implicit, causal assumptions (Uyeda  
43 et al. 2018) — and one must always keep in mind the motivating theory for how variables are functionally  
44 related to underlying causal processes and to assess whether we are indeed measuring the right thing (Houle  
45 et al. 2011). No matter how elegant the method, if it is not appropriate for the theoretical context, it is not  
46 going to tell you anything meaningful and even worse, will likely be misleading. We argue that inferences  
47 drawn in all comparative analyses should be carefully evaluated in light of their relationship to hypothesized  
48 causal relationships, and skeptically evaluated in light of the nature of our necessarily simplistic models.

49 Our motivating case study is a recent study by Avaria-Llautureo et al. (2019), which presents evidence  
50 from phylogenetic models that provides a new perspective on the major features of endothermic evolution,  
51 with the provocative conclusion that metabolic rate and body temperature are decoupled—a conclusion largely  
52 at odds with previous studies (Clarke et al. 2010). To support this conclusion, Avaria-Llautureo et al. (2019)  
53 use statistical phylogenetic comparative methods inspired by similar tests for convergence in selective pres-  
54 sures as demonstrated by convergent evolutionary rates, which are commonly used in molecular evolution  
55 (Chikina et al. 2016; Sackton et al. 2019). While superficially a reasonable analogy, we argue in this note  
56 that the novel conclusions of Avaria-Llautureo et al. (2019) are consequences of a failure to use models that  
57 reflect the processes that are important in the evolution of endothermic traits – and phenotypic traits more  
58 generally. By using process-based models and simulating different causal scenarios, we demonstrate how  
59 easily estimates from complex statistical models can become dissociated from the underlying biological hy-  
60 potheses. While our assumptions about cause-and-effect and/or biological constraints may be incorrect (e.g.  
61 if past processes are fundamentally different from extant processes), we argue it is far superior to make these  
62 assumptions explicit and part of the process of making inference from comparative data.

### 63 **Coupling of states or rates?**

64 It is well-established that the basal metabolic rate (*BMR*) of an organism is a function of its body mass  $M$ ,  
65 its internal body temperature  $T_b$ , and the temperature in which it lives (ambient temperature;  $T_a$ ) (Clarke  
66 et al. 2010, and references therein). These relationships form the basis of several ecological theories (e.g.,  
67 the Metabolic Theory of Ecology; (Brown et al. 2004)) and, more broadly, are used for ensuring that *BMR*

68 is measured in comparable way across individuals, species, and studies. That is to say,  $BMR$ ,  $M$ , and  $T_b$  are  
69 known to be functionally coupled. For this reason, the conclusion in [Avaria-Llautureo et al. \(2019\)](#) that there  
70 is no evidence of evolutionary or functional coupling between  $BMR$  and  $T_b$  may appear surprising.

71 However, here we look closer at how functional coupling can be defined and how it relates to the  
72 evolutionary questions being asked. In [Avaria-Llautureo et al. \(2019\)](#), they open their paper discussing this  
73 expected covariation of the values of  $BMR$ ,  $T_b$ , and  $T_a$ , but then propose defining “coupling“ to mean the  
74 correlation of evolutionary *rates* rather than *states*. This shift is subtle but important. Indeed, consistent with  
75 previous analyses of the same data, [Avaria-Llautureo et al. \(2019\)](#) do find evidence of a positive association  
76 between  $T_b$  and  $BMR$  ([Clarke et al. 2010](#)), and a negative association between  $T_b$  and  $T_a$  ([Clarke et al.](#)  
77 [2010](#)). We use this study as a motivating example to ask two questions that are important to consider when  
78 choosing from the variety of comparative methods available to test associations between traits. First, are the  
79 coupling of rates and the coupling of states simply alternative ways of measuring the same thing, or do they  
80 measure different things altogether? And second, if they are different, how does one choose a method that  
81 meaningfully corresponds to the theoretical context under study?

82 We illustrate our points in two ways. We first simulate trait data under three alternative and biologically  
83 plausible causal scenarios (depicted in Figure 1 and described below) on the phylogeny of mammals used  
84 by [Avaria-Llautureo et al. \(2019\)](#) and examine the resulting covariance structure in both the states and the  
85 rates. We then use the simulated scenarios as reference points for interpreting our re-analysis of the original  
86 empirical data of [Avaria-Llautureo et al. \(2019\)](#).

- 87 • **Scenario 1** is a multivariate Brownian motion (mvBM; [Felsenstein 1985](#)) model. Note that this model  
88 generates data with the same distribution as a Phylogenetic Generalized Least Squares (PGLS) model,  
89 though multivariate BM and PGLS depict different causal scenarios that are not identifiable in extant-  
90 only data ([Blomberg et al. 2012](#)). For simplicity, we simulate  $BMR$  and  $T_b$  under mvBM where the  
91 two traits are correlated ( $\rho = 0.8$ ) but where there is no variation in rates across different lineages  
92 (meaning there is no variation to detect a branch-wise correlation in rates).
- 93 • In **Scenario 2** we simulate uncorrelated states of  $T_b$  and  $BMR$  by changing the correlation in the  
94 evolutionary rate matrix to  $\rho = 0$ . However, we introduce correlations in branch-specific rates by  
95 drawing shared branch scalars for both traits from a  $\Gamma$  distribution (as is typical when capturing rate

96 variation in phylogenetic studies) with shape parameter set to 1.1 and a scale parameter set to 1. This  
97 results in collinear evolutionary rates for both traits, but no correlation in states. In other words, even  
98 though the traits are not correlated in any way, branches on which there is rapid evolution in one trait  
99 are also branches in which there is rapid evolution in the other trait. Perhaps surprisingly, we consider  
100 the most straightforward process consistent with this type of “rate-coupling” to be one in which there  
101 is a 3rd factor,  $Z$ , affecting both traits (Row 2, Figure 1). Should such a scenario be evidence of  
102 functionally relevant “coupling” as argued by [Avaria-Llautureo et al. \(2019\)](#)? We will return to this  
103 shortly.

- Finally, in **Scenario 3**, we first simulate  $T_b$  with branch scalars from  $\Gamma(shape = 10, rate = 10)$  and an overall Brownian Motion rate of  $\sigma_{T_b}^2 = 1$ . We then simulate another trait  $Z$  (which is not measured at the tips of the phylogeny) with branch scalars from the same  $\Gamma$ -distribution, but a higher rate of evolution where  $\sigma_Z^2 = 20$ . For each tip  $i$ , we then generate data on  $BMR$  by adding the effects of  $Z$  and  $T_b$ , such that

$$BMR_i = \beta_Z Z_i + \beta_{T_b} T_{b,i} + \epsilon_i$$

104 where  $\epsilon_i$  represents additional residual variation where  $\epsilon \sim \mathcal{N}(0, \sigma^2 = 1)$ . Because  $Z$  is unmeasured  
105 at the tips,  $Z$  is subsumed in the residual variation in the regression between  $BMR$  and  $T_b$ . Here  $Z$   
106 could represent body size or another related variable. Because the range of variation in  $Z$  is greater  
107 than  $T_b$ , we would expect that rate correlations between  $BMR$  and  $T_b$  would be difficult to detect even  
108 if there is a true underlying causal relationship, as is the case here (Row 3, Figure 1).

109 These simulated scenarios to guide our thinking on the meaning and relationships between rate and  
110 state associations. With these in mind, we re-analyzed the empirical datasets of [Avaria-Llautureo et al. \(2019\)](#)  
111 in two different ways. First, we fit a standard phylogenetic regression model ([Martins and Hansen 1997](#))  
112 to estimate a correlation between states (Row 4, Figure 1). We found that, as expected,  $T_b$  and  $\ln(BMR)$   
113 are positively correlated in both birds and mammals using models that include  $\ln(M)$  (and in the case of  
114 mammals,  $\ln(M)^2$ , since previous studies have found curvature in this relationship; [Kolokotronis et al.](#)  
115 [2010](#)). We found the best model for the residual variation by comparing Brownian Motion, Pagel’s  $\lambda$  ([Pagel](#)  
116 [1999](#)), and Ornstein-Uhlenbeck (OU; [Hansen 1997](#)) models with fixed or random roots fit using the R package  
117 `phylolm` ([Ho and Ané 2014](#)). In mammals, we found very strong coefficients between  $T_b$  and  $\ln(BMR)$

118 ( $\beta_{T_b} = 10\%$  increase in BMR per degree Celsius,  $P < 0.001$ ) and significant negative relationship between  
119  $T_a$  and  $T_b$  ( $\beta_{T_a} = -0.03^\circ C$  per  $^\circ C$  of  $T_a$ ,  $P < 0.01$ ). In birds, the former is marginally significant ( $P =$   
120  $0.057$ ) and also positive ( $\beta_{T_b} = 2\%$  increase in BMR per degree Celsius), and the latter is non-significant  
121 ( $P = 0.45$ ). We note that very little variation exists for avian  $T_b$  and that proper analysis of these correlations  
122 would likely require careful accounting of measurement error (Hansen and Bartoszek 2012). These results  
123 are consistent with previous analyses of these relationships (Clarke et al. 2010).

124 Second, we estimate the evolutionary correlation in rates. We note that rate correlations are consider-  
125 ably more difficult to estimate than state correlations since we cannot actually measure these in any organism  
126 and must infer them from model parameters; furthermore, even perfectly-estimated rates carry only infor-  
127 mation about the magnitude — not the direction — of change. In their analyses, Avaria-Llautureo et al.  
128 (2019) estimated branch specific rates using a Variable-Rate Regression model (Venditti et al. 2011) fit using  
129 Bayesian Reversible Jump MCMC (Green 1995). The model they fit is quite complex; as such, in order to  
130 visualize the essence of their results without getting mired in the details (e.g., choice of priors, assessment  
131 of convergence, etc.) we plot the signal for rate correlations in branch-specific rates using the logarithm of  
132 the absolute value of independent contrasts computed at each node (Garland et al. 1992). While an imper-  
133 fect measure, the expected value of each contrast will increase with increasing evolutionary rate and thus  
134 we can test whether these node-wise estimates of rates are correlated between different sets of traits. For  
135  $\ln(BMR)$ , we again took the residuals of the best-fitting phylogenetic regression with log body mass (and  
136 for mammals,  $\ln(M)^2$ ) to calculate these contrasts. To verify that correlated rate scalars can be identified  
137 using this approach, we simulated scenarios of rates with shared shift locations on the mammal phylogeny  
138 with correlations in rates ranging from 0 to 1 and evaluated whether significantly positive regressions were  
139 obtained (see Supplementary Material on Dryad, <https://doi.org/10.5061/dryad.z612jm6bj>).

140 For mammals, there is weak but marginally significant evidence that  $\ln(BMR)$  and  $T_b$  have correlated  
141 evolutionary rates ( $\rho = 0.10$ ,  $P = 0.03 - 0.08$ , depending on how contrasts of 0 are treated; Fig. 1.4C).  
142 Our simulations indicate that this value of the slope is consistent with the conclusion of a weak to moderate  
143 correlation between rates (Supplementary Material). For birds, there is no apparent relationship ( $\rho = 0.04$ ,  
144  $P = 0.58$ , Fig. 1.4C). Both of these results are qualitatively in-line with the findings of Avaria-Llautureo et al.  
145 (2019) and we do not dispute their results in this regard. Instead, we wish to highlight the importance of the  
146 interpretation of rate correlations, rather than to dispute the methods themselves. Rather, we argue that the

147 presence or absence of associations of rates can occur with or without functional coupling, and care should  
148 be taken in their interpretation. For metabolic rate and body temperature, we consider that the combination  
149 of strong state coupling and weakly-detectable rate-coupling using contrasts, and the variable-rate regression  
150 model in [Avaria-Llautureo et al. \(2019\)](#), to be consistent with our *Scenario 3*. We therefore read the evidence  
151 to suggest that  $T_b$  is functionally coupled with *BMR*, but that metabolic rate is also strongly affected by other  
152 traits with additional, independent rate variation.

153 Furthermore, our *Scenario 2* clearly illustrates that shared rate variation can exist even if there is no  
154 functional or evolutionary association between the traits (cf. [Avaria-Llautureo et al. \(2019\)](#)'s definition of  
155 “coupling”; their Fig. 1b, our Figure 1.2A-C). For example, consider if both traits were evolving with rates  
156 jointly set by a third factor ( $Z$ ). This could occur if rates of change in traits evolving via genetic drift are all  
157 jointly set by effective population size ( $Z = N_e$ ; [Walsh and Lynch 2018](#)), even when traits are functionally  
158 uncoupled. Thus, we observe that strong and easily detectable causal relationships between traits can be  
159 found by state associations (as we observed in the empirical data), even when coupling of rates are too weak  
160 to be detectable (Figure 1). Furthermore, tests of rate coupling can produce positive results even when traits  
161 are not interdependent — which seems to us the most relevant questions to the evolutionary and ecological  
162 theories that motivate interest in these traits — but instead whether an additional causal factor might be  
163 driving shared rate variation among the two traits.

164 It is worth noting that a number of researchers investigating molecular evolution have interpreted  
165 “convergent rates” in molecular sequence evolution as possible evidence of shared functional significance  
166 ([Hu et al. 2019](#); [Smith et al. 2020](#)). Rather than assessing convergent states (e.g. [Li et al. \(2010\)](#)), such studies  
167 ask whether shared selective pressures cause joint shifts in substitution rates across genes—which corresponds  
168 closely to our *Scenario 2*. While not the focus of the present manuscript, we think it is worth thinking  
169 carefully about the underlying logic of such tests for convergence in molecular evolution. Importantly, studies  
170 examining genomic characters have the advantage that additional data is often available to support claims of  
171 evolutionary coupling, such as the enrichment for meaningful functional annotations against a large sample  
172 of background rates and *a priori* hypotheses of specific causal factors ([Chikina et al. 2016](#); [Sackton et al.](#)  
173 [2019](#)). While these additional data make evaluating shared function via coupled rates a more meaningful  
174 inference, we would strongly caution against interpreting a lack of evidence for joint shifts in substitution  
175 rates as evidence of an absence of functional relationship between molecular sequences—consistent with the

176 arguments we have made here.

177         Given the existing evidence and known mechanism for how  $T_b$  and  $BMR$  are coupled, combined  
178 with the ease of testing state relationships with continuously-varying traits (as opposed to the complexities  
179 of evaluating convergent states in molecular sequences), the evidence and test for functional coupling here is  
180 much more direct. In the context of metabolism, we believe that subsequent evaluation of *Scenario 2* in this  
181 case, at least without reference to a background suite of traits or a specific causal factor, is largely meaningless.  
182 We acknowledge that we may not have fully understood the causal model envisioned by [Avaria-Llautureo  
183 et al. \(2019\)](#). For this reason, we think it is critical that practitioners of phylogenetic comparative methods  
184 present graphical causal models of their hypotheses to make clear how the inferences drawn relate to the  
185 evidence presented.

## 186 **Coupled trends or model inadequacy?**

187 Thinking clearly about the traits under study also requires we consider the causal processes that affect their  
188 evolution. This choice of process model can critically affect how we interpret the data, and helps explain  
189 another somewhat surprising feature of metabolic evolution found by [Avaria-Llautureo et al. \(2019\)](#): they find  
190 that  $T_b$  has *decreased* consistently throughout the Cenozoic. Of course, the vast amount of evidence from both  
191 physiology and paleontology strongly supports the conclusions that endotherms originated from ectotherms  
192 ([Grigg et al. 2004](#); [Lovegrove 2017](#)) — this statement is not controversial and it would be extraordinary if  
193 this was found to be incorrect. Nevertheless, a plausible alternative is that [Avaria-Llautureo et al. \(2019\)](#)  
194 identified a signal of endotherms rapidly increasing to high average body temperatures and subsequently  
195 decreasing over the Cenozoic.

196         In order to evaluate these alternatives, we first recognize that all models are simplifications of reality,  
197 and necessarily make simplifying assumptions. Despite this caveat, we believe that flawed models can often  
198 be used to make robust macroevolutionary inferences. We argue that the key to progress despite imperfect  
199 tools is to consider carefully how such traits are *expected* to evolve given our knowledge of their function, dy-  
200 namics and constraints – knowledge that comes from outside of macroevolutionary statistical models. Here,  
201 we re-examine the conclusions of [Avaria-Llautureo et al. \(2019\)](#) to demonstrate our view of this process, and  
202 how our expectations can easily explain the observed patterns with entirely expected violations of standard

**Table 1:** Phylogenetic signal estimated using an OU model of adaptive (i.e. constrained) evolution.

Clade	Trait	Phylogenetic half-life <sup>†</sup>	$\Delta AIC$ over BM
Mammals			
	$T_b$	0.26	46.3
	$BMR^{\ddagger}$	0.16	86.0
	$T_a$	0.10	141.2
Birds			
	$T_b$	0.01	132.4
	$BMR^{\ddagger}$	0.01	382.4
	$T_a$	0.01	168.8

<sup>†</sup> Half-life ( $\frac{\ln(2)}{\alpha}$ ) measures the time it takes for a lineage to evolve half-way to the OU optimum in units of tree height. Values of 0 indicate constrained evolution with no phylogenetic signal; while values >1 indicate BM-like evolution and strong phylogenetic signal. All traits strongly reject the BM model ( $\Delta AIC > 4$ ).

<sup>‡</sup> Mass-corrected log BMR.

203 macroevolutionary models.

204 As stated above, [Avaria-Llautureo et al. \(2019\)](#) used a Variable Rates Regression model based on a  
205 Brownian motion model of evolution, where traits evolve in an un-directed and unbounded manner as they  
206 would under genetic drift or randomly varying selection ([Hansen and Martins 1996](#)). This seems entirely  
207 inappropriate for traits such as *BMR* that are biophysically constrained to stay close to an optimum value ([West  
208 et al. 2002](#); [Glazier 2005, 2010](#); [Clarke et al. 2010](#)). While far from perfect, the “adaptation” (or OU) model  
209 proposed by [Hansen \(1997\)](#) better captures this biological reality. Analyzing the data of [Avaria-Llautureo  
210 et al. \(2019\)](#) bears this out: OU is a much better fit than Brownian motion for  $T_b$ ,  $T_a$  and mass-specific  
211 *BMR* in both mammals and birds (Table 1). Birds are especially strongly constrained with little variation in  
212 these traits and virtually no phylogenetic signal; this is consistent with both the basic physiology involved as  
213 well as with previous phylogenetic analyses ([Uyeda et al. 2017](#)). We note that the models we used for this  
214 comparison are much simpler than the ones used by [Avaria-Llautureo et al. \(2019\)](#); comparing variable OU  
215 processes ([Uyeda et al. 2017](#)) with variable BM processes is beyond the scope of this paper, but we argue  
216 that the simple case captures the essence of the problem.

217 So how could the choice of Brownian models over constraint models affect the apparent trend in  $T_b$ ?  
218 Let’s assume for the moment that there is no rate variation in the data and that the evolutionary dynamics of  
219  $T_b$  and  $T_a$  do indeed resemble that of an OU process. One feature of evolutionary processes that follow an  
220 OU model is that since the amount of divergence is constrained, the overall rate one would measure if one  
221 fits a Brownian motion model would depend strongly on the length of the branch itself (the denominator of

222 any estimate of rate); therefore if one uses a Brownian model to fit data generated under an OU-like process,  
223 rates will be estimated to be higher on short terminal branches, whereas deeper and longer branches will have  
224 lower estimates of rate (for mathematical explanation, see [Uyeda et al. 2015](#)). If one computes, as [Avaria-  
225 Llautureo et al. \(2019\)](#) did, the total pathwise rates (summing the rates on all the branches from the root to a  
226 given tip), one will detect highest pathwise rates in any trait that is correlated with short terminal branches.  
227 [Avaria-Llautureo et al. \(2019\)](#) interpret correlations between pathwise rates and the tip state to be evidence  
228 of a secular evolutionary trend. But given the likely dependence on the rate estimates for  $T_b$  and  $T_a$  with  
229 branch lengths, any association between these traits and diversification will result in longer pathwise-rates.  
230 For example, there is abundant evidence that terminal branches are shorter in temperate regions than in the  
231 tropics for these taxa owing to higher species turnover in temperate regions ([Weir and Schluter 2007](#); [Schluter  
232 2016](#); [Schluter and Pennell 2017](#)). Thus, we think a far more likely explanation is that this trend is an artifact  
233 of fitting Brownian models to constrained data.

234 Indeed, we are able to recover the exact same pattern of decreasing  $T_b$  over time as [Avaria-Llautureo  
235 et al. \(2019\)](#) find in their Fig. 4, even when we simulate a true trend of *increasing*  $T_b$  over time. For exam-  
236 ple, starting from a low ancestral body temperature of (25°C), it is straightforward to obtain negative rela-  
237 tionships between pathwise-rates and body temperature under biologically realistic macroevolutionary land-  
238 scapes (Fig. 2). Specifically, we used a constrained macroevolutionary landscape that reflects standard enzy-  
239 matic and temperature preference curves for organisms in which high temperatures impose a stronger con-  
240 straint on organismal performance than low temperatures (i.e., the Sharpe-Schoolfield equation [Schoolfield  
241 et al. 1981](#), see supplement for details of simulations). In other words, this model is similar to an OU model  
242 in that divergence is constrained within high-performance regions of the macroevolutionary landscape, but  
243 allows asymmetry that is expected given well-known physiological performance curves for  $T_b$  ([Huey and  
244 Kingsolver 1989](#)). We simulated data on this landscape with the R package [BBMV \(Boucher et al. 2017\)](#).  
245 These asymmetries are expected to generate a negative relationship between  $T_b$  and the pathwise-rates of  $T_b$   
246 *even if there is a true trend towards higher body temperatures towards the present*. This also could explain  
247 why [Avaria-Llautureo et al. \(2019\)](#) conclude that  $T_a$  and  $T_b$  follow the same trajectory toward decreasing tem-  
248 perature over time (*cf.* Fig. 1.2A-C), despite evidence they are negatively correlated with each other when  
249 we consider state-correlations ([Clarke et al. 2010](#))—which likely results from the well-known phenomenon of  
250 counter-gradient selection ([Schultz et al. 1996](#); [Fangue et al. 2009](#)).

251 While estimation of ancestral states under a VRRM may be possible if the trait approaches uncon-  
252 strained BM-like evolution, we caution interpretation of ancestral states even in the best circumstances —  
253 and especially with evidence of constrained evolution and when trends are expected. The inability to estimate  
254 the ancestral states for constrained traits is a special case of the “Darwinian uncertainty principle” (Gascuel  
255 and Steel 2019), which describes the trade-off in estimating ancestral states vs. the rates of the evolutionary  
256 process. Just as in the case of molecular sequence data, quantitative traits with high rates that explore a con-  
257 strained macroevolutionary landscapes will progressively erase evolutionary history, eroding any confidence  
258 we should have in already uncertain ancestral state estimates Boucher et al. (2017).

## 259 Conclusions

260 Phylogenetic comparative datasets cannot be generated from experimentation, and as such causal claims are  
261 viewed with appropriate skepticism. However, it is vital to not throw away our causal and process-based  
262 evolutionary thinking when applying the increasingly complex and sophisticated statistical toolkit of com-  
263 parative methods. Instead, our recommendation is to make explicit the causal reasoning that justifies the use  
264 of a particular macroevolutionary model – even when such models are likely gross simplifications of real-  
265 ity. Furthermore, we recommend critically evaluating the conclusions of that model in light of likely model  
266 violations from the known biology of the trait, even when evaluating these assumptions may be impossi-  
267 ble. In our specific example, we examined the relationships between metabolic rate and temperature. While  
268 these relationships have been studied by physiologists for decades, we certainly think there is potential for  
269 novel phylogenetic comparative methods to provide new insights into the problem (White et al. 2009; Uyeda  
270 et al. 2017; White et al. 2019). However, the causal assumptions of inferences drawn from statistical meth-  
271 ods, which are often implicit, must be consistent with the fundamental biology underlying the data (Uyeda  
272 et al. 2018). This is especially true since complex phylogenetic comparative models can have many-to-one  
273 mappings of interpretations to patterns (Louca and Pennell 2020). Realizing the potentially large extent of  
274 such issues in macroevolution, we believe that it is vital to carefully consider how plausible biological causal  
275 processes *would* map onto macroevolutionary patterns, even when definitively inferring causation from ob-  
276 servational data alone is impossible. This step we think is too often ignored in macroevolutionary models,  
277 and by no means unique to the question and study examined here. We hope that our exploration has pro-

278 vided a useful model for how we can make not just our methods and data analyses reproducible, but also our  
279 reasoning and inferences.

280 We demonstrate in this note that we are at odds with the interpretation found in [Avaria-Llatureo et al.](#)  
281 (2019), who concluded that 1) *BMR* and  $T_b$  are evolutionarily decoupled; 2) this decoupling is likely related  
282 to positive correlations between cooling ambient global temperatures and decreasing  $T_b$ ; 3) and that  $T_b$  and  
283  $T_a$  have decreased since the origin of endothermy. We present evidence that these three major claims rely on,  
284 in our opinion, a flawed mapping of statistical methods to relevant causal processes and fitting models that  
285 are fundamentally inconsistent with the biological context. By applying (admittedly, also flawed) models that  
286 capture key components of the underlying process and its constraints, we conclude that 1)  $T_b$  and *BMR* are  
287 strongly coupled in evolutionary state and possibly weakly in evolutionary rate 2)  $T_b$  is negatively correlated  
288 to  $T_a$  in mammals and 3) that evidence for a decrease in  $T_b$  over the course of endotherm evolutionary  
289 history could likely be a spurious result of model inadequacy and the constraints imposed by physiology.  
290 More broadly, we want to encourage researchers to recognize the limitations of phylogenetic comparative  
291 models and think critically about how to model well-studied biological processes. That is to say, we must  
292 choose our statistical tools based on biology rather than let our view of biology be shaped by our choice in  
293 statistical tools.

## 294 **Acknowledgements**

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## 298 **Conflicts of Interest**

299 The authors have no conflicts of interest to declare.

## Data Availability

Supplementary Material and complete scripts and data are available on Dryad, <https://doi.org/10.5061/dryad.z612jm6bj>.

## Author contributions

JCU and MWP conceived of the manuscript. JCU, NB, SM performed analyses. JCU, NB, SM, JR, and MWP wrote the manuscript.

**Figure 1:** Simulated (Rows 1-3) and Empirical (Row 4) relationships between  $T_b$  and BMR on the mammalian phylogeny used in Avaria-Llautureo et al. (2019). (Row 1; *Scenario 1*): If  $T_b$  is direct cause of BMR or if both evolve under a common cause,  $Z$ , in a correlated fashion (1A), then we observe a correlation in state (1B; dotted line = PGLS regression) despite an absence of rate variation (1C) [cf. Avaria-Llautureo et al. (2019) Fig.1a]. (Row 2; *Scenario 2*): Multivariate Brownian motion (2A) where  $T_b$  and BMR are uncorrelated in state (2B), but share the same source of rate variation (2C). Shared rate variation indicated by colored nodes/arrows. This is the scenario that Avaria-Llautureo et al. (2019) define as evolutionary “coupling” [cf. Avaria-Llautureo et al. (2019) Fig.1b], despite the fact that it can occur absent a functional relationship between traits (e.g., evolution by genetic drift with  $Z$  representing time-varying effective population size). (Row 3; *Scenario 3*)  $T_b$  is a direct cause of BMR and has rate variation, but  $Z$  is also a cause of BMR with independent rate variation (3A). Rates in  $Z$  are 20x the rates in  $T_b$ . This results in highly significantly correlated states (3B), but non-significantly correlated rates (3C) [cf. Avaria-Llautureo et al. (2019) Fig.1c]. Plotted rate scalars for simulated data are true parameter values, giving a best case scenario. Using estimated rate scalars would drastically decrease power to detect significant rate correlations even further; particularly for methods that “shrink” rate variation (like the variable rates regression model – VRRM). However, state correlations will remain robustly estimated. (Row 4): Empirical data for birds and mammals used in Avaria-Llautureo et al. (2019) using phylogenetically mass-corrected log BMR. Rates are estimated as the log of magnitude of the phylogenetic independent contrasts. P-values from full, best-fitting phylogenetic regression models. The empirical data is consistent with the causal model in (3A), showing strong evidence of “evolutionary coupling” between  $T_b$  and BMR. Details and complete R script are available on Dryad; <https://doi.org/10.5061/dryad.z612jm6bj>.

**Figure 2:** Relationship between pathwise rates estimated from (A) empirical data [cf. Avaria-Llautureo et al. (2019) Fig. 4] and (B) 10 simulated datasets with increasing  $T_b$  over time. This negative slope was used by Avaria-Llautureo et al. (2019) to support their conclusion that  $T_b$  and  $T_a$  had decreased over the evolutionary history of mammals and birds. Analyses of data with the VRRM in *bayestraits* (green) and in *bayou* (blue) are shown with dotted lines indicating the phylogenetic regression. When the true simulated process includes constraints (instead of purely unconstrained BM) under realistic macroevolutionary landscapes, we recover negative relationships between pathwise-rates and trait values, despite an overall increasing trend (from the root value of 25°C to an optimum of 38°C). This macroevolutionary landscape was represented by the Sharpe-Schoolfield thermal-performance curve (gray dotted line), which has a harder bound at high temperatures than at low temperatures. Pathwise-rates should never be interpreted as reconstructing ancestral states when the trait in question shows evidence of strongly constrained evolution, as constrained evolution will erase phylogenetic signal and measured rates will simply either reflect asymmetries in the macroevolutionary landscape (as depicted here) and/or asymmetries in diversification/taxonomic sampling (e.g. trait-dependent diversification, likely the case for  $T_a$ ). Data were simulated with the R package *BBMV* Boucher et al. (2017). Supplementary Material and complete scripts and data are available on Dryad, <https://doi.org/10.5061/dryad.z612jm6bj>.

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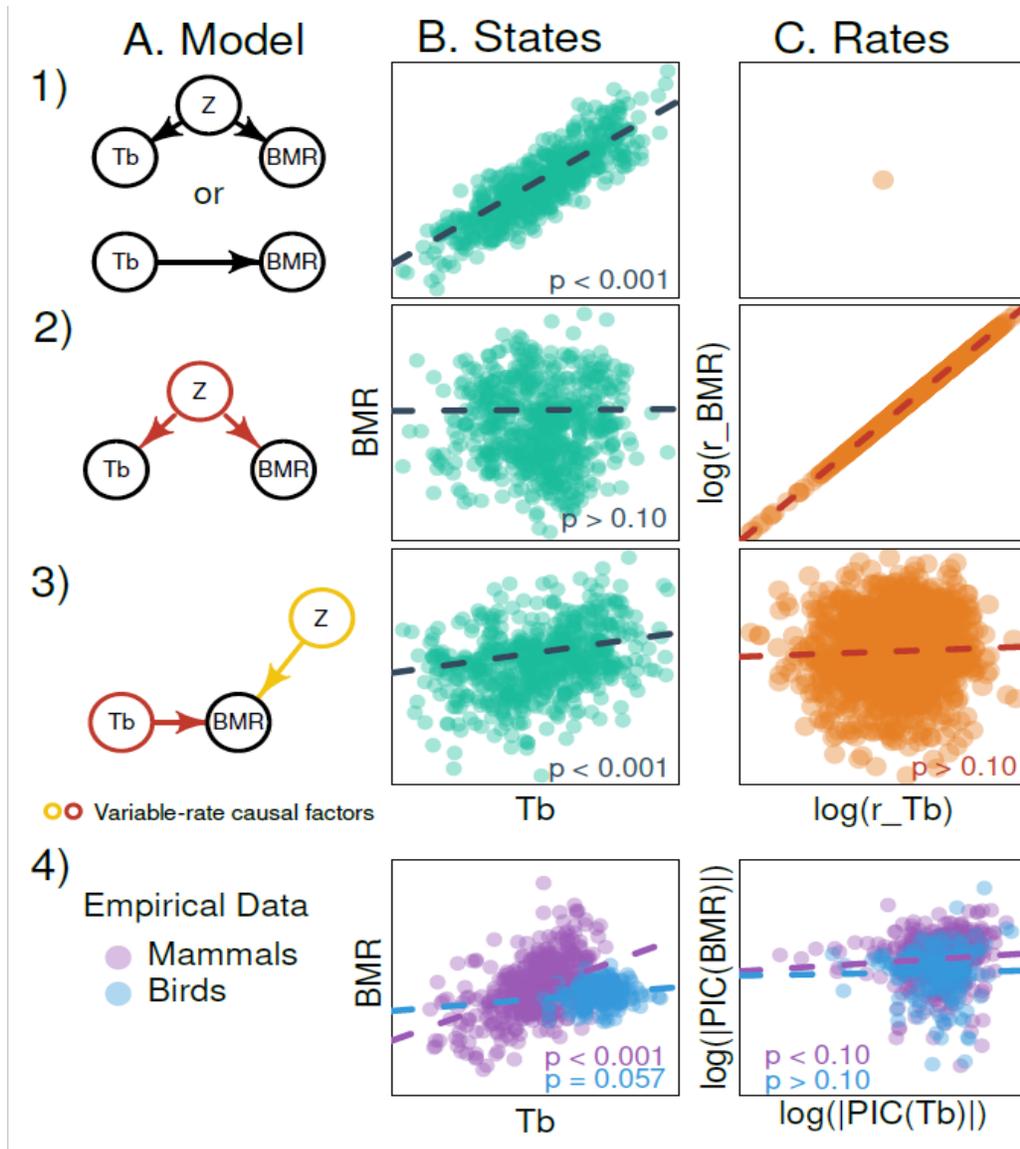
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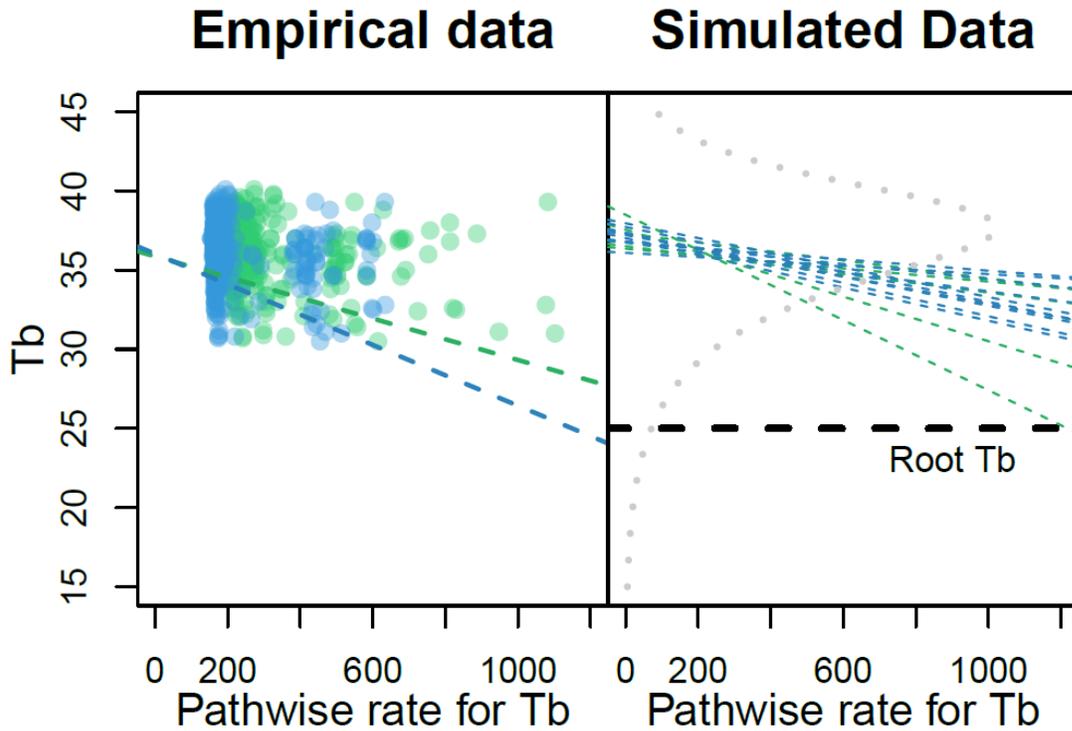
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